Emerging signals modulating potential of ginseng and its active compounds focusing on neurodegenerative diseases

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Abstract
Common features of neurodegenerative diseases (NDDs) include progressive dysfunctions and neuronal injuries leading to deterioration in normal brain functions. At present, ginseng is one of the most frequently used natural products. Its use has a long history as a cure for various diseases because its extracts and active compounds exhibit several pharmacological properties against several disorders. However, the pathophysiology of NDDs is not fully clear, but researchers have found that various ion channels and specific signaling pathways might have contributed to the disease pathogenesis. Apart from the different pharmacological potentials, ginseng and its active compounds modulate various ion channels and specific molecular signaling pathways related to the nervous system. Here, we discuss the signal modulating potential of ginseng and its active compounds mainly focusing on those relevant to NDDs.

1. Introduction

In the nervous system, neurodegenerative diseases (NDDs) are multifactorial debilitating disorders that are characterized by progressive dysfunction and neuronal injury leading to a slow but irreversible deterioration of brain functions which affects around 30 million individuals worldwide [1–4]. Although some symptomatic treatments are available, specific treatments have not yet been discovered [5]. Ginseng is considered to be one of the widely used traditional herbal medicines [6]. Ginseng and its constituents have been documented to produce aphrodisiac, adaptogenic, immunomodulatory, antiinflammatory, antioxidant, antiaging, anticancer, antifatigue, antidiabetic, pulmonary protective, hepatoprotective, cardioprotective, hypolipidemic, and renoprotective effects in various studies [7–21].

Furthermore, ginseng extracts and its active compounds have exhibited properties including antistress, antidepressive, and neuroprotective in various studies of neurological disease models [22–30]. They also enhanced cognitive performance and help maintain brain health [30–34]. Recently, various studies have reported on the potential role of ginseng and its active compounds in treating neurological disorders. Here, focusing on potential therapies for NDDs, we present the signal modulating potential of ginseng and its active compounds.

2. Role of ginseng and its active compounds in modulating ion channel signaling pathways

2.1. Modulation of voltage-gated ion channel

Various sources of evidence have suggested that ginsenosides regulate the neuronal Na⁺ channels. In Xenopus oocytes, ginsenoside Rg3 carbohydrate component inhibited the inward Na⁺ peak current [35]. Also, in Xenopus oocytes, Rg3 inhibited the Na⁺ channel by acting on the resting and open states of the Na⁺ channel via contact with the S4 voltage sensor segment of domain II [36]. Moreover, ginsenoside Rh2 inhibited the Na⁺ channel function by inhibiting veratridine-dependent depolarization of mouse synaptoneurosomes following the inhibition of l-glutamate and γ-aminobutyric acid releases [37].

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By activating the K⁺ channel, some ginsenosides regulated the electrical state of excitatory neurons. In *Xenopus* oocytes, ginsenoside Rg3 enhanced outward large-conductance Ca²⁺ and voltage-gated big K⁺ (BKCa) channel currents in a concentration-dependent, voltage-dependent and reversible manner [38]. Moreover, in rat’s brain, ginsenoside Rf-activated G protein-gated inwardly rectifying potassium (GIRK) channels through an unidentified G protein-coupled receptor [39].

In neuronal cells, ginsenosides are capable of inhibiting the Ca²⁺ channels. Ginseng total saponins are the main bioactive ingredients of *P. ginseng*. By inhibiting L-type Ca²⁺ channels, the ginseng total saponins decreased the KCl-induced neuronal loss in primary cortical neurons [40]. In Xenopus oocytes, BKCa channels are heterologously expressed. Gintomin treatment activated the BKCa channel and expressed the α-subunit of the BKCa channel in a concentration-dependent manner [41]. In primary hippocampal neurons, through L-type Ca²⁺ channels, ginsenoside Rg1 inhibited Ca²⁺ influx [42]. Furthermore, in the amyloid beta (Aβ)25–35 model, ginsenoside Rg1 inhibits high-voltage-activated calcium channel currents in hippocampal neurons [43]. In addition, ginsenoside Rb1 inhibited voltage-gated calcium channel currents in a concentration-dependent manner, and upon washout, the current was mostly recovered. In hippocampal neurons, Rb1 selectively inhibits the action of L-type voltage-gated calcium channels without affecting the N-type or P/Q-type Ca²⁺ channels [44]. The signal modulating effects of active ginseng compounds on voltage-gated ion channels are shown in Fig. 1.

### 2.2. Modulation of the ligand-gated ion channel

#### 2.2.1. Nicotinic acetylcholine receptors

Active ginseng compounds modulate nicotinic acetylcholine receptors (nAChRs). In oocytes expressing nAChR subunits (α1β1δε or α3β4), ginsenosides inhibited the acetylcholine (ACh)-induced inward currents (I_ACh). This potential indicates that ginsenosides directly control the nAChR channel activities. In the case of I_ACh inhibition, protopanaxatriol (PPT) ginsenosides (Re, Rf, Rg1, or Rg2) were more powerful than protopanaxadiol (PPD) ginsenosides including Rb1, Rb2, Rg1, and Rd [45]. Conversely, by the desensitization of ACh induced in oocytes expression, ginsenoside Rg2 reduced the peak current and elevated the inward currents in human nAChR subunits α3β4, α3β2, α4β4, and α4β2 [46]. Moreover, through nAChR channel activity, ginsenosides show the inhibitory effects on I_ACh reduction of catecholamine release. Regarding the heterologous expression of the α9x10 subunits of nAChR in *Xenopus* oocytes, I_ACh is blocked by ginsenosides with a potency order of Rg3 > Rb2 > CK > Re ≈ Rg2 > Rf > Rc > Rb1 > Rg1 in a resolvable means. Ginsenoside Rg3’s blocking effects on I_ACh were the same after preapplication linked to ginsenoside Rg3 co-application. Furthermore, α10 subunit of α9x10 nAChR might play an important role in Rg3-induced regulation of the α9x10 nAChR [47]. Besides, a recent study described the ameliorative effect of Rg1 on lipopolysaccharide (LPS)-induced cognitive deficits. Rg1 treatment inhibited LPS-induced reduction of ACh levels and an increase in acetylcholinesterase activity. LPS treatment reduced the α7 nAChR protein expression in the prefrontal cortex and hippocampus, but Rg1 treatment reverted the changes [48]. Furthermore, choline acetyltransferase (ChAT) and vesicular acetylcholine transporter (VACHT) are essential for cholinergic neurotransmission. Ginsenosides Rg3, Rg2, and Re regulated both ChAT and VACHT. In Neuro-2a cells, the Rg and Re effectively induced the ChAT/VACHT genes expression and ACh promotion [49].

#### 2.2.2. γ-amino butyric acid receptors

Ginsenosides interact with the γ-amino butyric acid (GABA_B) receptor and might regulate the ligand binding with the GABA_B receptor. In a rat brain membrane fraction, ginsenosides differentially regulate the binding of [3H]-flunitrazepam or [3H]-muscimol to the GABA_B receptor [50]. Conversely, longer infusion of ginseng and ACh promotion [49].
regulatory effects of ginsenoside metabolites differ from those of ginsenosides. The human recombinant GABA<sub>α</sub> receptor (α<sub>1</sub>β<sub>2</sub>γ<sub>2</sub>) channel activity expressed in Xenopus oocytes, M4, a metabolite of PPT ginsenosides, more potently inhibited the I<sub>GABA</sub> than PPD. The effect of M4 and PPD on I<sub>GABA</sub> was both concentration-dependent and reversible. The half-inhibitory concentration (IC<sub>50</sub>) values of M4 and PPD were 17.1 ± 2.2 and 23.1 ± 8.6 μM, respectively. The inhibition of I<sub>GABA</sub> by M4 and PPD was voltage-independent and noncompetitive [53]. Using a conventional whole-cell patch-clamp technique in acutely isolated rat hippocampal CA3 pyramidal neurons, compound K produced a potential effect on GABAergic spontaneous miniature inhibitory postsynaptic currents. Compound K increases spontaneous GABA release by increasing intraterminal Ca<sup>2+</sup> stores [54]. A recent report indicated that GABA<sub>α</sub> receptor agonist pretreatment significantly potentiated the Panax quinquefolius neuroprotective effect. Regarding the sleep deprivation, GABAergic mechanism induced anxiety-like behavior, mitochondrial dysfunction, oxidative stress, hypothalamic–pituitary–adrenal axis activation and neuroinflammation that could be involved in the P. quinquefolius neuroprotective outcome [55].

2.2.3. Glutamate receptors channel activity

In different neurotoxic agents–induced models, active ginseng compounds produced effects that confirmed the potential effects of ginseng active compounds on glutamate receptors. In rat hippocampal cultures, Rg3 reduced the high K<sup>+</sup>- activated glutamate, and N-methyl-D-aspartate (NMDA)-induced Ca<sup>2+</sup> influx significantly [56]. Ginseng total saponins decreased glutamate-induced cultured rat astrocytes swelling [57]. Ginsenosides Rb1 and Rg3 produced a protective effect against glutamate-induced neurotoxicity. In this study, Rb1 and Rg3 prevent the nitric oxide (NO) overproduction, malondialdehyde formation. The treatments also preserved the superoxide dismutase level and diminished the Ca<sup>2+</sup> influx in rat cortical cultures [58]. In the Huntington’s disease model, the neuroprotective effects of ginsenosides Rb1, Rc, and Rg5 might be due to the inhibition of glutamate-induced Ca<sup>2+</sup> responses in cultured medium spiny striatal neuronal culture [59]. In cultured mouse cortical neurons, notoginsenoside R1 (NGR1) also prevented glutamate-mediated neurotoxicity [60]. Pretreatment with ginsenosides attenuated NMDA- or substance P-induced nociceptive behavior through the intrathecal route [61,62]. With respect to the ginsenosides, the pretreatment attenuates the kainate-induced cellular death in hippocampal neurons [63]. Ginsenoside Rb3 could exert a neuroprotective role on hippocampal neurons, a role which was partly mediated by the facilitating Ca<sup>2+</sup>-dependent deactivation of NMDA receptors and the resulting reduction of intracellular free Ca<sup>2+</sup> level [64]. Ginsenosides Rh2 and Rg3 prevent the NMDA receptor channel currents in cultured rat hippocampal neurons [65]. Moreover, in vitro and in vivo studies of homocysteine (HC)-induced hippocampal excitotoxicity, ginsenoside Rg3 produced neuroprotective activity. Furthermore, in vivo experiments showed that intracerebroventricular Rg3 preadministration significantly and dose-dependently decreased the HC-induced hippocampal damage in rats. Treatment with Rg3 has been found to dose-dependently inhibit the HC-induced elevation of intracellular Ca<sup>2+</sup> levels. In addition, in Xenopus oocytes expressing the NMDA receptor, Rg3 treatment dose-dependently repressed HC-induced currents [66]. Regarding the effects of ginsenoside metabolites on NMDA receptor channel activity, PPT contrasting compound K and PPD were reversibly repressed the NMDA-mediated inward currents (I<sub>NMDA</sub>) in a dose-dependent manner. I<sub>NMDA</sub> inhibition by PPT was noncompetitive with NMDA and was self-regulating the membrane holding potential, although ginsenoside Rh2, Rg3, and PPT interrelate with the NMDA receptor [67]. Ginsenoside Rb1 produced the anti-fatigue effect through the inflammatory cytokine-mediated NMDA receptor pathway [68]. The protective effects of active ginseng compounds on toxins-induced excitotoxicity through the glutamate receptors are shown in Fig. 2.

3. Various specific molecular signaling and their modulating effect by ginseng and its active compounds

3.1. Toll-like receptor involving pathways

In an Alzheimer’s disease (AD) cellular model, ginsenoside Rg1 produced antineuroinflammatory activity through a toll-like receptor (TLR) pathway. In NG108-15 cells, Aβ<sub>25−35</sub> markedly

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Fig. 2. Potential activities of active ginseng compounds in toxin-induced neurotoxicity models. GTS, ginseng total saponins; NGR1, notoginsenoside R1. Glutamate, homocysteine and kainate cause an increase in intracellular calcium level leading to excitotoxicity. Active ginseng compounds produce protective activities against toxins-induced excitotoxicity.
increased the expressions of TLR3 and TLR4. In a concentration-dependent manner, Rg1 significantly reduced the expressions of both proteins as well as mRNA of TLR3 and TLR4 [69].

3.1.1. Mitogen-activated protein kinase pathway

Red ginseng marc oil (RMO) reduced the p38 mitogen-activated protein kinase (MAPK) and its upstream kinases including MAPK kinases 3/6 (MKK3/6) phosphorylation. By blocking the p38 MAPK pathway, RMO produced an antiinflammatory effect in LPS-induced RAW 264.7 macrophages [70]. Ginseng pectin pretreatment enhanced the phosphorylation of both the extracellular signal-regulated kinases 1 and 2 (ERK1/2) in cortical neuron cells and in U87 cells, it also increased the ERK1/2 phosphorylation. Owing to the activation of the phosphorylation of ERK1/2, ginseng pectin produced this neuroprotective activity against H2O2-induced apoptosis [71]. In LPS- induced RAW 264.7 cells, synthesized gold nanoparticles (AuNPs) using Panax ginseng extract antiinflammatory effects through p38 MAPK [72]. Likewise, gintonin produced antiinflammatory activity via the signal transduction through MAPK, and it potently suppressed the NO production and also efficiently suppressed the proinflammatory cytokines levels in RAW 264.7 cells [73].

Different doses of ginsenoside Rb1 pretreatment noticeably attenuated tau protein hyperphosphorylation and the expression of c-Jun N-terminal kinase (JNK)/p38 MAPK in Aβ25-35-induced model [74]. In 1-methyl-4-phenylpyridinium (MPP+)−treated PC12 cells, Rb1 improved ERK1/2 phosphorylation and reduced phosphorylated p38 or stress activated protein kinase/JNK. Rb1 increased ERK1/2 phosphorylation, and it was abrogated by estrogen receptor siRNA [75]. By upregulating the growth-associated protein 43 (GAP-43) expression through ERK-dependent signaling pathways, ginsenoside Rb1 may help PC12 cells neutre outgrowth [76]. In addition, Rb1 efficiently inhibited the activation of the MAPK signaling pathway induced by spinal cord injury in the rat model, which might be involved in the neuroprotective activity of Rb against spinal cord injury [77].

In various studies, ginsenosides Rg1 and Rg5 have shown potential effects through this pathway. To promote cell proliferation, ginseng and Rg1 increase the MAPK signaling pathways. In RSC96 cells, ginseng and Rg1 were recognized to have proliferative effects that are MAPK signaling-dependent [78]. In addition, Rg1 produced antiinflammatory activity in BV-2 microglial cells. In this study, phosphoinositide phospholipase C-γ1 inhibition was moderately abolished, and Rg1 produced an inhibitory effect on ERK1/2, JNK, and p38 MAPK phosphorylation. Therefore, by suppressing the neurotoxic proinflammatory mediators and cytokines expression, Rg1 expressively attenuates the overactivation of microglial cells through phospholipase C-γ1 signaling pathway activation [79]. Besides, Rg1 improves the neurite outgrowth and defends against Aβ25-35− induced damage, and this mechanism may be involved in the activation of ERK1/2 signaling [80]. Additionally, Rg1 activated the ERK/MAPK pathway in another study [81]. Ginsenoside Rg5 inhibited the phosphorylation of MAPKs and the DNA binding activities in LPS-stimulated BV-2 microglial cells and primary rat microglia [82].

NGR1 and compound K show potential activity through the modulation of this pathway. NGR1 produces a sufficient effect by inducing an estrogen receptor−dependent ERK1/2 pathway [83]. Compound K significantly suppressed the Phorbol-12-myristate-13-acetate−mediated p38 MAPK, ERK, and JNK activation, which are upstream modulators of activator protein−1. In addition, compound K also inhibited the invasiveness of in vitro glioma cells [84].

3.1.2. Nuclear factor kappa-light-chain-enhancer of activated B cells pathway

RMO and AuNPs obtained from the leaf extract of P. ginseng and gintonin produced antiinflammatory activities in the LPS-induced RAW 264.7 macrophage cell line [70,72,73]. RMO treatment reduced the inducible nitric oxide species and cyclooxygenase-2 at both the mRNA and protein levels, with a blockade of the nuclear translocation of the p65 subunit. Hence, due to the inhibition of nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) transcriptional activity, RMO might produce this antiinflammatory effect [70]. Moreover, AuNPs decreased inflammatory mediators, including NO, prostaglandin E2, interleukin-6, and tumor necrosis factor−α, expression. In RAW 264.7 cells, AuNPs suppressed the LPS-induced activation of NF-κB and COX-2 at the mRNA and protein levels, with a blockade of the nuclear translocation of the p65 subunit. Hence, due to the inhibition of nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) transcriptional activity, RMO might produce this antiinflammatory effect [70]. Moreover, AuNPs decreased inflammatory mediators, including NO, prostaglandin E2, interleukin-6, and tumor necrosis factor−α, expression. In RAW 264.7 cells, AuNPs suppressed the LPS-induced activation of NF-κB and COX-2 at the mRNA and protein levels, with a blockade of the nuclear translocation of the p65 subunit. Hence, due to the inhibition of nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) transcriptional activity, RMO might produce this antiinflammatory effect [70]. Moreover, AuNPs decreased inflammatory mediators, including NO, prostaglandin E2, interleukin-6, and tumor necrosis factor−α, expression. In RAW 264.7 cells, AuNPs suppressed the LPS-induced activation of NF-κB and COX-2 at the mRNA and protein levels, with a blockade of the nuclear translocation of the p65 subunit. Hence, due to the inhibition of nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) transcriptional activity, RMO might produce this antiinflammatory effect [70]. Moreover, AuNPs decreased inflammatory mediators, including NO, prostaglandin E2, interleukin-6, and tumor necrosis factor−α, expression. In RAW 264.7 cells, AuNPs suppressed the LPS-induced activation of NF-κB and COX-2 at the mRNA and protein levels, with a blockade of the nuclear translocation of the p65 subunit. Hence, due to the inhibition of nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) transcriptional activity, RMO might produce this antiinflammatory effect [70]. Moreover, AuNPs decreased inflammatory mediators, including NO, prostaglandin E2, interleukin-6, and tumor necrosis factor−α, expression. In RAW 264.7 cells, AuNPs suppressed the LPS-induced activation of NF-κB and COX-2 at the mRNA and protein levels, with a blockade of the nuclear translocation of the p65 subunit. Hence, due to the inhibition of nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) transcriptional activity, RMO might produce this antiinflammatory effect [70].

In the AD model, P. ginseng ginsenosides Rg1, Rg3, Rd, and Rg3 enriched the extract, producing potential activities [83−88]. In an advanced glycation end product−induced AD model, ginseng showed the neuroprotective effects through the significant decrease in the expression of the receptor for advanced glycation end-products and NF-κB in a rat [86]. In transgenic mice, Rd might improve learning and memory ability in used Aβ1 precursor by inhibiting the transcription activity of NF-κB. Moreover, by suppressing the activated NF-κB pathway, the proinflammatory cytokines were further reduced, and the protective factors were generated [87]. In a scopolamine-induced model, the oral administration of ginsenoside Rg3-enriched ginseng extract (Rg3GE) suppressed the increase in acetylcholinesterase activity and stimulation of the NF-κB pathway (i.e., phosphorylation of p65) in the hippocampus [88]. In the LPS-induced BV-2 microglia cell line, ginsenosides Re and Rh1 produced antiinflammatory activity [89,90]. Re produced neuroprotective events through phospho-p38, iNOS, and COX-2 signaling pathways [89]. Without affecting NF-κB DNA binding, ginsenoside Rh1 inhibited LPS-induced NF-κB−mediated transcription. In addition, an increase in cAMP responsive element-binding protein was identified to result in the suppression of NF-κB−mediated transcription [90]. Moreover, in a study of oxidative stress, ginsenoside Rg1 normalized the oxidative stress−induced nonmuscle myosin heavy chain IIA (NMHC II A) overexpression in PC12 cells. The collected data showed that the NMHC II A/NF-kappa B/p65 pathway was involved in oxidative stress−induced cell death [91]. The effects of active ginseng compounds through the MAPK and NF-κB pathways are summarized in Table 1.

3.2. Caspase-3 and Bcl-2-like protein 4-mediated pathways

Ginsenosides showed potential activity through these pathways. NGR1 produced neuroprotective activity by suppressing caspase-3 activation [83]. In PC12 cells, ginsenoside Rb1 prevented MPP+−induced caspase-3 activation and DNA fragmentation. Besides, Rb1 also activated B-cell lymphoma−extra-large (Bcl-xL) and reduced apoptosis [75]. Ginsenoside Rg1 inhibited the caspase-3 signaling pathway and myosin IIα−actin interaction, and through these inhibitions, it produced neuroprotective activity [92]. In an MPP+−induced apoptosis in human neuroblastoma (SH-SY5Y) cells model, Rg1 can effectively decrease the expression of MPP+−induced upregulation of Bax and decrease the B-cell lymphoma 2 (Bcl-2) expressions. In addition, Rg1 can effectively decrease the MPP+−induced toxicity by inhibiting the activation of caspase-3 [93]. A recent study revealed that ginsenoside Rd prevents tri-methyltin-induced cell apoptosis via regulation of caspase-3, Bcl-2, and Bcl-2−like protein 4 [94]. With respect to the ginsenoside Rb1, it suppressed the activation of ER stress−associated proteins including protein kinase RNA−like endoplasmic reticulum kinase
and C/EBP homology protein and high glucose induced Bcl-2 downregulation in hippocampal neurons [95].

### 3.3. Phosphoinositide 3-kinase/protein kinase B pathway

Ginseng protein produced neuroprotective activity in D-galactose/AlCl₃-induced AD model, and protective effect is connected to the phosphoinositide 3-kinase/protein kinase B (PI3K/Akt) signaling pathway [96]. Ginseng protein decreased the Aβ₁₋₄₂ content and phosphor-tau and enhanced the mRNA and PI3K and phospho-Akt/Akt protein expression in the hippocampus [96]. Ginsenoside Rd promotes the glutamate clearance, which was achieved by upregulating the glutamate transporter 1 expression via the PI3K/Akt pathway [97]. Neurite outgrowth is a crucial process associated with neuronal repair. In addition, Rd produced an effect on neurite outgrowth, and the upregulation of GAP-43 expressions through PI3K-Akt-dependent pathways might be responsible for this activity [76]. Furthermore, an experimental Parkinson’s disease model induced by MPP⁺ in SH-SY5Y showed that different concentrations of Rd had a neuroprotective effect. This protective effect might be due to the PI3K/Akt survival-signaling pathway [98]. Various studies have shown that ginsenosides Rg1 and Rg5 show beneficial effects through this pathway. Ginsenoside Rg1 enhances

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<th>Ginseng(active compounds)</th>
<th>Models</th>
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<td>Rh1</td>
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<td>Different doses attenuate tau protein hyperphosphorylation and the expression of JNK/p38 MAPK in the process.</td>
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<td>MPP⁺-treated PC12 cells</td>
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<td>Rd</td>
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<td>Spinal cord injury in rat</td>
<td>Produces neuroprotective activity by efficiently inhibiting the activation of the MAPK signaling pathway.</td>
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<td>APP transgenic mice</td>
<td>By inhibiting the transcription activity of NF-κB, might improve learning and memory when APP is used. Moreover, by suppressing the activated NF-κB pathway, further reduces the pro-inflammatory cytokines and generates protective factors.</td>
<td>[87]</td>
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<td>Rg1</td>
<td>Aβ₁₋₄₂-induced NG108-15 cells RSC96 cells</td>
<td>Reduces the increased expressions of both TRL3 and TLR4.</td>
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<td>Aβ₁₋₄₂-induced cultured hippocampal neurons PC12 cells</td>
<td>Attenuates the overactivation of phosphoinositide phospholipase C-γ1 and produces the inhibitory effect on the ERK1/2, JNK and p38 MAPK phosphorylation.</td>
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<td>H₂O₂-induced PC12 cells</td>
<td>Improves neurite outgrowth and defends against damage, and the mechanism may comprise the activation of ERK1/2 signaling.</td>
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<td>By CaMKIIα, it activates the ERK/MAPK pathway.</td>
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<td>Normalizes the oxidative stress-induced NMMCH IIA overexpression. Regarding the collected data, NMMHC IIA-NF-κappa B/p65 pathway involved in oxidative stress-induced cell death.</td>
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<td>Rg3GE</td>
<td>Scopolamine-induced mice</td>
<td>Suggests the increase in acetylcholinesterase activity and stimulation of the NF-κB pathway (i.e., phosphorylation of p65) in the hippocampus.</td>
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<td>Rg5</td>
<td>LPS-stimulated BV-2 microglial cells and rat primary microglia</td>
<td>Inhibits the phosphorylation of MAPKs and the DNA binding activities.</td>
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<td>Ginseng Pectin</td>
<td>H₂O₂-induced apoptosis in cortical neuron cells and U87 cells</td>
<td>Pretreatment enhances the phosphorylation of both the extracellular signal-regulated kinases 1 and 2 (ERK1/2).</td>
<td>[71]</td>
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<td>AuNPs</td>
<td>LPS-induced RAW 264.7 cells</td>
<td>Exerts anti-inflammatory effects through the suppression of NF-κB signaling pathway activation through p38 MAPK.</td>
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<td>LPS-induced RAW 264.7</td>
<td>Effectively suppresses the NO production without any cytotoxicity and also proficiently suppresses the proinflammatory cytokines levels. Furthermore, facilitates signal transduction through NF-κB pathways and recovers the mir-34a and mir-93 levels.</td>
<td>[73]</td>
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<td></td>
<td>H₂O₂-induced PC12 cells</td>
<td>Produces neuroprotective activity the effect by inducing an estrogen receptor-dependent ERK1/2 pathway.</td>
<td>[83]</td>
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<td>Compound K</td>
<td>Phorbol myristate acetate—mediated human astrogliaoma cells</td>
<td>Significantly suppresses the p38 MAPK, ERK, and JNK activation, which are upstream modulators of activator protein-1.</td>
<td>[84]</td>
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<td>Ginseng</td>
<td>Advanced glycation end product—induced AD in rat</td>
<td>Shows neuroprotective effects through the significant decreasing the expression of receptor for advanced glycation end-products and NF-κB.</td>
<td>[86]</td>
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</table>

AD, Alzheimer’s disease; APP, amyloid β-protein precursor; AuNPs, synthesized gold nanoparticles; CaMKIIα, calcium/calmodulin-dependent protein kinase type II alpha chain; JNK, c-Jun N-terminal kinase; LPS, lipopolysaccharide; MAPK, mitogen-activated protein kinase; NF-κB, nuclear factor kappa-light-chain-enhancer of activated; NGR1: notoginsenoside R1; NMMCH IIA, nonmuscle myosin heavy chain IIA; NO, nitric oxide; Rg3GE, Rg3-enriched ginseng extract; RMO, Red ginseng marc oil; SAPK, stress activated protein kinase; TLR, toll-like receptor.
neurite outgrowth and protects against Aβ25–35-induced damage, and its mechanism may involve the activation of Akt signaling [71]. In LPS-stimulated BV-2 microglial cells and rat primary microglia, Rg5 produced an anti-inflammatory activity. The studies indicated that Rg5 inhibited the phosphorylation of PI3K which are upstream molecules controlling the inflammatory reactions [82]. Moreover, pseudoginsenoside-F11 produced antineuroinflammatory activity on activated microglia. Pseudoginsenoside-F11 inhibited neuroinflammation LPS-induced in N9 microglia by inhibiting the activation of TLR4 mediated PI3K/Akt pathways [99]. NGR1 produced a neuroprotective activity by suppressing the H2O2-induced intracellular reactive oxygen species accumulation and increasing the product of lipid peroxidation (malondialdehyde), protein oxidation (protein carbonyl), DNA fragmentation (8-OHdG), mitochondrial membrane depolarization as well as caspase-3 activation. This neuroprotective activity is occurred by inducing estrogen receptor-dependent crosstalk between Akt pathways [83].

3.4. Insulin-like growth factor-I receptor signaling

A study of MPP+-induced apoptosis model in PC12 cells showed that ginsenoside Rb1 had neuroprotective effects. In caspase-3-dependent apoptosis pathway, Rb1 protects the PC12 cells through the estrogen receptor [75]. In a study on RSC96 Schwann cells, ginseng and ginsenoside Rg1 exhibited proliferation and migration-enhancing properties. Ginseng and ginsenoside Rg1 improve the insulin-like growth factor-I receptor (IGF-IR) pathway regulators' protein expression. Moreover, Rg1 with biomedical materials would be a possible method to improve neuron regeneration [78]. Ginsenoside Rg1 produced neuroprotective effects against 6-hydroxydopamine (6-OHDA)—induced neurotoxicity [100,101]. In the 6-OHDA—induced model of nigrostriatal injury, Rg1 showed a neuroprotective activity, and this effect might involve the activation of the IGF-IR signaling pathway. Rg1 treatment ameliorated this behavior in apomorphine-induced rotational behavior. In addition, 6-OHDA significantly reduced the striatum dopamine content, and Rg1 reversed the significant effects of 6-OHDA [101]. Besides, in 6-OHDA—induced neuronal damages in SK-N-SH cells, Rg1 produced neuroprotective effects through the activation of the IGF-IR, ER signaling pathways, and its antiapoptotic potentials [100].

3.5. Nuclear factor (erythroid-derived 2)-like-2 factor pathway

P. ginseng extract produced antidepressant activity in a chronic restraint stress-induced depression model in mice. In this study, P. ginseng extract showed anti-neuroinflammatory and antioxidant, nuclear factor (erythroid-derived 2)-like-2 factor/heme oxygenase-1 (Nrf2/HO-1 activation) activity by inhibiting the hypothalamo—pituitary—adrenal axis mechanism [102]. Ginsenoside Rb1 activates the Nrf2/HO-1 signaling pathway to display a potent neuroprotective activity against tert-butyldihydroperoxide—induced oxidative injury. In cultured neural progenitor cells, Rb1 activates the Nrf2 pathway and led to an elevated HO-1 expression [103,104]. In iron-induced neurotoxicity, the antioxidative properties that trigger the Akt/Nrf2 pathway and increase the Nrf2-induced HO-1 and Cu/Zn superoxide dismutase expression allowed ginsenoside Rg1 protect the human neuroblastoma SK-N-SH cells [105]. Similarly, ginsenoside Rg3 improves the Nrf2 DNA-binding activity [106]. PPT ginsenosides improve the Nrf2 inflowing to the nucleus and induced antioxidant response elements (ARE) such as HO-1 and the expression of nicotinamide adenine dinucleotide phosphate (NAPDH) quinone oxidase 1. Subsequently, PPT shows a neuroprotective activity against 3-nitropipionic acid—induced injury (oxidative stress) in the rat model of Huntington's disease [107]. Additionally, NGR1 improves Nrf2 nuclear translocation and ARE activity. It also upregulates the expression and effects of HO-1, NADPH quinone oxidase 1, and gamma-glutamylcysteine synthetase. NGR1 provides neuroprotective activity by inducing the estrogen receptor-dependent activating Nrf2/ARE signaling [83].

3.6. Nerve growth factor and brain-derived neurotrophic factor pathways

Various research studies have shown that ginseng extracts, as well as its active compounds, exert potential activity through these pathways. In Neuro-2a cells, ginsenosides Rd and Re treatments meaningfully improved the microtubule-associated protein-2, nerve growth factor receptor (p75), p21, and tropomyosin receptor kinase A genes and proteins expression. Therefore, Re and Rd play a significant role in differentiation of neuron and the nerve growth factor—tropomyosin receptor kinase A signaling pathway [49].

In the rat model, P. ginseng showed neuroprotective activity against LPS-induced brain injury. LPS causes elevated brain NO and serum HC associated with a reduction of brain-derived neurotrophic factor (BDNF) level. On the other hand, in the treated group, P. ginseng significantly attenuated these compared to the LPS group [108]. A study showed that P. notoginseng saponins promote the rat embryonic cortical neural stem cells survival, self-renewal, proliferation, and differentiation through neurotrophic factors in the autocrine or paracrine signaling [109].

Ginsenoside Rg1 also improved the memory performance [110,111]. In the AD model, Rg1 treatment reduced the Aβ1-42 accumulations and phosphorylated (p)-Tau protein. Rg1 treatment also upregulated the BDNF and phosphorylated tropomyosin receptor kinase B. Similarly, Rg1 treatment also restored the long-term potentiation in the AD mice model. In another study, through the hippocampal BDNF upregulation, Rg1 treatment improved the memory performance in middle-aged mice, changing apical spines and helping hippocampal long-term potentiation [111].

Ginsenoside Rb1 revealed a neuroprotective activity against cerebral ischemia. After Rb1 infusion, the number of nestin-positive cells apparently increased. At different points in time, Rb1-treated rats showed a BDNF level that significantly improved compared to that of ischemic rats. These neuroprotective effects might be because of the promotion of the neurogenesis and expression of BDNF regulation [112]. In addition, Rb1 showed the preventive and therapeutic effects on the neural injury during cerebral infarction in rat’s model via middle cerebral artery occlusion. The increases in occlusion duration resulted in a decline in interleukin-1 levels and GAP-43 level and an increase in BDNF levels and neurofilaments. These effects might be due to a decrease in inflammation and assistance in the growth of nerve cells [113].

Ginsenosides Rb3, Rg1, and Rg3 produced antidepressant activity through the BDNF signaling pathway [114–117]. In a study of the chronic mild stress model, treatment with Rb3 significantly increased the BDNF level in the prefrontal cortex and hippocampus area [114]. Rg1 produced antidepressant activity via BDNF signaling pathway. It upregulates the BDNF expression and hippocampal neurogenesis [115,116]. Recently, Rg1 treatment prevented chronic social defeat stress—induced depressive-like symptoms in a depression-induced mouse model [116]. Furthermore, Rg3 completely restored the chronic social defeat stress—induced reduction in the hippocampal BDNF signaling pathway [117].

3.7. Mechanistic target of rapamycin signaling, Wnt/β-catenin and Rho-associated kinase 1/Myosin light-chain kinase pathways

Compound K is produced potential activity through the mechanistic target of rapamycin pathway. In a study of AD model, compound K promotes Aβ1 clearance and enhances autophagy in
primary astrocytes. It also slows AD pathological progression. In addition, due to the mechanistic target of rapamycin phosphorylation, it might contribute to an enhancement in the autophagy [118]. In vivo and in vitro Parkinson’s disease models, ginsenoside Rg1 showed neuroprotective potentials through the Wnt/b-catenin signaling pathway. In both the in vivo and in vitro studies, Rg1 showed protective effects on the protein and mRNA expression levels of this pathway marker. With respect to the in vitro study, the neuroprotective potentials were blocked by Dickkopf-related protein 1 [119]. In a H2O2-induced model study, treatment with Rg1 eliminated H2O2-induced different morphological fluctuations such as cell rounding, membrane blebbing, neurite retraction, and nuclei condensation. The mechanism of neuroprotection of Rg1 is connected to the inhibition of myosin IIa–actin interaction and the signaling pathway of Rho-associated kinase 1/Myosin light-chain kinase [92].

4. Conclusion

Recently, medicinal plants have shown potent pharmacological roles in various disorders that have been proven in several pre-clinical and clinical studies. Identifying the proper therapeutic targets is essential to setting up treatment strategies for various neurological disorders. In case of NDDS, no cure has yet been discovered, and this is a major topic of research today. In addition, identifying therapeutic targets and therapeutic molecules are crucial steps for the designing of therapeutic management for NDDS. Considering the importance of having targets, several signals have demonstrated a potential pharmacological role in various neurological disorders. Detecting these targets, allows the possibility to discover novel compounds suitable to treat the NDDS. Ginseng and its active substances have been known as multipotential therapeutics against various acute and chronic diseases. With respect to their multipotential role, researchers have concentrated their research. Various reports have indicated that they may be great therapeutic agents for neurological disorders due to their potential activities that modulate the various molecular signaling pathways.

However, research has not concluded how ginseng active compounds can have specific medicinal effects on NDDS. We focused on NDDS and here present the ion channels and specific molecular signals that are modulated by ginseng and its active substances. The overview shows that ginseng components might emerge as candidates for NDDS due to their versatile potential, specifically, their neuroprotective activities against neuroinflammation and apoptotic cell death. It might be helpful to conduct deeper research and clinical trials regarding NDDS. Also, based on signal modulating potential, molecular modification of ginseng compounds may be useful in obtaining superior pharmaceutical drugs. In addition, understanding of the complex response to ginseng could allow developing synergistic drug therapy.

To examine and validate the potential, several disease models were designed including toxin-induced cellular and animal models as well as transgenic models to conduct for different NDDS. Further studies concerning specific ion channels and molecular signals known in the pathogenesis and as therapeutic targets for NDDS are suggested to confirm the exact pharmacological and therapeutic activities of ginseng and its active compounds. The design and execution of clinical studies for active ginseng compounds to treat NDDS is a major challenge for the clinical scientist. Before clinical studies, more investigations using laboratory-based and informatics-based approaches should be conducted to understand the precise medicinal efficacy, potency, and safety in the NDDS. These approaches might give direction to modify and synthesize new derivatives of active ginseng compounds targeting the NDDS.

Finally, the diverse potential role of ginseng active compounds might serve a good role in treatment strategies to cure NDDS. In near future, rigorous studies of active ginseng compounds with their modified forms will produce drugs for therapy of NDDS.

Conflicts of interest

The authors have no conflicts of interest.

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